

Original Research Article

<https://doi.org/10.20546/ijcmas.2017.607.278>

Cultural and Physiological Studies on Wild Mushroom Specimens of *Schizophyllum commune* and *Lentinula edodes*

B. Praveen Kumar Reddy¹, A. Uma Rajashekhar², P. Harikrishna^{3*} and A.V.N. Lavanya⁴

¹Department of Horticulture, College of Horticulture, Dr. Y.S.R. Horticultural University, Rajendranagar, Hyderabad, 500030, Andhra Pradesh, India

²Department of Agriculture Microbiology and Bio-energy, ANGRAU, College of Agriculture, Rajendranagar, Andhra Pradesh, India

³Department of Agriculture Microbiology and Bio-energy, ANGRAU, Post-Harvest Technology Centre, College of Agriculture, Bapatla, Andhra Pradesh, India

⁴Department of Horticulture, College of Horticulture, Dr. Y.S.R. Horticultural University, Venkataramannagudem, Andhra Pradesh, India

*Corresponding author

ABSTRACT

Keywords

Schizophyllum commune,
Lentinula edodes,
Physiological
studies, Culture
media.

Article Info

Accepted:
23 June 2017
Available Online:
10 July 2017

A study was conducted on the influence of culture media, temperature and pH levels on the mycelial growth of two wild specimens of *Schizophyllum commune* and *Lentinula edodes* during 2013. The results of the experiment indicated that, potato dextrose agar medium with a pH range of 5.0-7.0 at a temperature of 30°C and malt extract medium with a pH range of 4.5-6.5 at a temperature of 25°C were found optimum for the mycelial growth of *S. commune* and *L. edodes*, respectively. The sharp decline in the pH values of the liquid broth media under post mycelial growth conditions of *L. edodes* clearly indicated the prefer ability of the media or substrates with lower pH. Study of lethal temperatures revealed the tropical nature of the wild strain of *S. commune*, where there was revival of the growth even after the exposure period of seven days at 45 °C. The results will be helpful for the maintenance of these cultures in pure, stable and viable conditions.

Introduction

Mushrooms are increasingly being evaluated around the world for their nutritional and pharmacological properties. They are considered as functional foods as well as a source of physiologically beneficial and non-invasive medicines. Since the ancient times medicinal mushrooms have been played significant role in alleviating the health issues

of human beings (Neha and Bharat, 2012). The traditional use of mushroom products has been long established among different cultural groups.

As a result of the large scale domestication of these nutritionally and medicinally important mushrooms, mushroom production became

the second most important commercial microbial technology, next to the yeasts (Singh *et al.*, 2000).

Different mushrooms were studied by the scientific community, in searching for new therapeutic alternatives. Some of the most recently isolated compounds from the shiitake (Martnexplores *et al.*, 2009) and *Schizophyllum* (Arpita and Bhupendra, 2013) have shown promising immunomodulatory and anti-tumor effects. Presently, the interest in the use of these mushrooms as a functional food is rising, but not much advancement has been made due to the availability of less number of strains suitable to grow under controlled conditions and also lack of pure and viable cultures for spawn production at a larger scale. Periodic collections of indigenous mushroom specimens from various climatic zones of the country and their cultural and morphological characterization are necessary to avert this problem and also for the future exploitation of the beneficial compounds from the wild mushrooms. Sizable production of these mushrooms depends upon proper maintenance and production of reliable pure culture under stable and viable conditions. The morphological conditions for the most efficient mycelial colonization of the wild specimens need to be standardized for their use as reference strains. Hence, the present investigation was undertaken to determine the physiological factors required for the conservation and maintenance of pure cultures of the wild specimens of *Schizophyllum commune* and *Lentinula edodes*.

Materials and Methods

The wild specimens of *Schizophyllum commune* and *Lentinula edodes* were collected in the Plant Pathology Department, College of Agriculture, Mushroom

Cultivation Scheme, Rajendranagar, Hyderabad, India and the tissue culture of the specimens were raised from sporophores on MEA medium. These specimens were purified and cultures were deposited in the Microbiology lab at Department of Microbiology, Post-Harvest Technology Centre, College of Agriculture, Bapatla, Andhra Pradesh, India for conservation and coded as SC-1 (*Schizophyllum commune*) and LE-1 (*Lentinula edodes*).

Various physiological factors viz., media, temperature and pH required for optimal vegetative growth and conservation of the cultures were evaluated. Malt Extract Agar (MEA), Potato Dextrose Agar (PDA), Potato Malt Agar (PMA), Potato Carrot Agar (PCA), Oat Meal Agar (OMA) and Raper's Complete medium (RC) containing 20.0 g glucose, 2.0 g peptone, 2.0 g yeast extract, 0.50 g magnesium sulphate, 0.46 g potassium dihydrogen phosphate and 20.0 g agar powder per liter of distilled water were used in the study as mycological media after sterilized in an autoclave at 121⁰ C and 15 psi pressure for 20 minutes.

The effect of different temperatures on mycelial growth of both the wild specimens was studied by inoculating the mycelial discs of similar diameter (8 mm). The discs were obtained by using the sterilized punch-hole tool to cut the mycelium from the periphery of 10 days old pure culture and place it into a new sterilized medium plate by using a transplant needle. Plates were incubated separately at temperatures of 10-40 °C with an interval of 5 °C and the radial growth was recorded at 24 hrs intervals.

To find out the optimum pH level, mycelia dry weight was recorded as the measure of biomass production in malt extract broth (malt extract powder 30.0 g, peptone 5.0 g per liter of distilled water). The pH of media was

adjusted from 3.5-9.0 (each at an interval of 0.5) by using N/10 NaOH or N/10 HCl. Microscopic studies of the vegetative mycelium were also undertaken to study the morphological characteristics of vegetative mycelium.

The treatments were arranged in a Completely Randomized Design (CRD) with five replications for each treatment and the data on experimental observations were statistically analyzed by using software AGRES.

Results and Discussion

Effect of different media on mycelial growth rate (mm/day)

Both the isolates could grow on MEA, PDA, PMA, PCA and RC medium. However, SC-1 showed significantly higher growth rate of 17.00 mm per day on PDA medium and LE-1 showed an average growth of 12.32 mm per day on MEA media which was significantly higher compared to the other mycological media (Table 1). Evaluation of culture conditions for the vegetative growth of different strains of *L. edodes* by Hem Lata and Sharma, 2012 also revealed significantly higher mycelial growth rate of 7.5 mm/day on MEA media. Least growth rate of 6.05 and 2.73 mm per day was observed in OMA media for SC-1 and LE-1 respectively. The pattern of mycelial growth of both the wild strains on PDA, PMA and MEA in petri plates were fully compact whereas, fluffy and sparse growth was observed on PCA, OMA and RC medium. Microscopic studies of the vegetative mycelium clearly showed the clamp connections at each septum which may be helpful for the future taxonomic studies of these wild specimens.

Effect of pH on mycelial growth

pH of the medium is important as it influences the mineral availability and

enzyme activity. It is evident from the data that, pH range of 5.0 to 8.0 was optimum for the growth of *S. commune*, whereas pH range of 4.5 to 6.5 was found best for the optimum growth of *L. edodes* (Fig. 1). pH range of 4.0-7.0 was recorded optimum for the vegetative growth of *L. edodes* in the earlier studies (Tnglet *et al.*, 2006). The exact hydrogen environment of fungi is difficult to study as fungi changes the pH when it grows on the media. The critical observation of pH values of the filtrate after fungal growth explains the above pattern.

During the mycelial growth on liquid broth media the strain SC-1 modified the pH of the media to the nearest optimum range from 5.47 to 6.58 (Fig. 2) which supported the best growth of the strain on the above mentioned range. The pH values of the liquid broth media after filtration of the biomass produced by the strain LE-1 showed an extreme acidic range of 3.11 to 3.38 (Fig. 1) which indicates the prefer ability of acidic substrates for the vegetative growth of *L. edodes*. Correlation studies of the *L. edodes* crop performance with the substrate properties also showed the sharp decline in the pH values of the substrate after incubation period in the earlier studies (Philippoussis *et al.*, 2003).

Effect of temperature on mycelial growth rate (mm/day)

The results indicated the typical tropical nature of the *S. commune* which has shown considerable measure of mycelial growth at a wide range of temperatures. It was found that the growth rate of *S. commune* was optimum (15 mm per day) at the temperature of 30 °C and the mycelia continues to grow (1.7 mm per day) even at 40 °C.

The growth rate was revived to 12.86 mm per day when the strain was shifted to the optimum temperature after 10 days exposure at 40 °C (Table 2). Based on the experimental

findings, 25 °C was found optimum temperature for the culture maintenance of *L. edodes* where the growth rate was observed @ 8.45 mm per day. Studies conducted on

Malaysian strain of *L. edodes* showed the best growth rate of 12.85 mm/day at a temperature of 25 °C (Sharma *et al.*, 2006) which confirms the present findings.

Fig.1 Biomass production (Dry weight in mg) at different pH levels

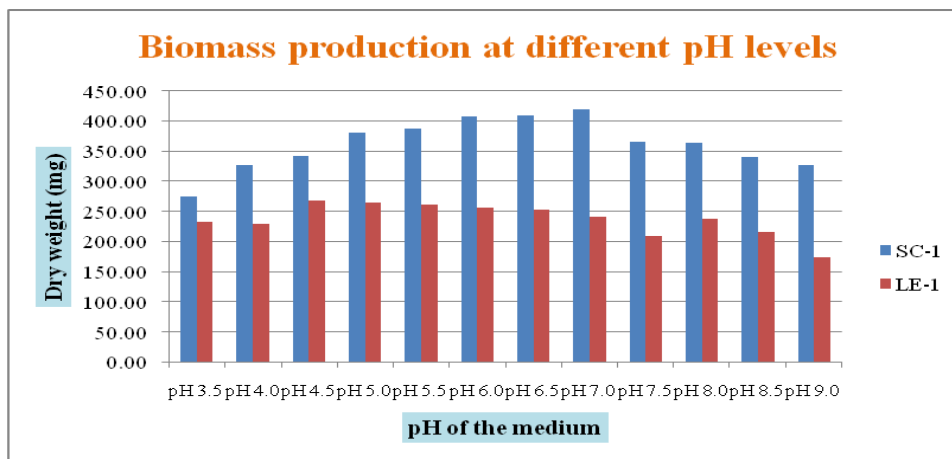


Fig.2 Post filtration pH values of liquid broth media

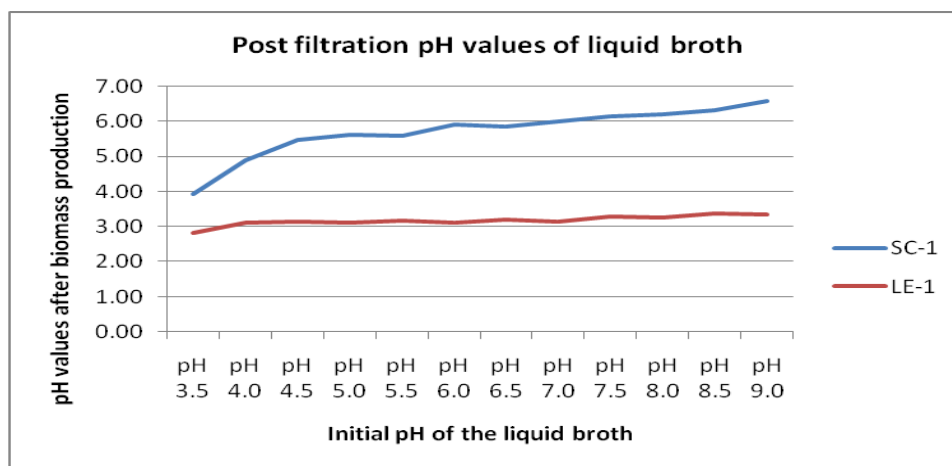


Table.1 Effect of different culture media on mycelial growth rate (mm/day)

Culture medium	Radial mycelial growth (mm/day)	
	SC-1	LE-1
Oat Meal Agar	6.05	2.73
Potato Dextrose Agar	17.00	9.67
Raper's Complete medium	14.28	5.30
Potato Malt Agar	14.28	9.33
Malt Extract Agar	14.28	12.32
Potato Carrot Agar	11.78	10.00
S Ed	0.98	0.45
CD (0.05)	2.98	1.39

Table.2 Effect of different temperatures on mycelial growth rate (mm/day)

Temperature	Radial mycelial growth (mm/day)	
	SC-1	LE-1
10 °C	2.31	2.06
15 °C	2.43	2.43
20 °C	9.67	6.29
25 °C	11.25	8.45
30 °C	15.00	5.31
35 °C	13.57	No growth
40 °C	5.51	No growth
SE d	0.43	0.21
CD (0.05)	1.27	0.94

Table.2a Lethal temperatures for the mycelial growth (mm/day)

Strain	Temperature	Exposure interval		
		7 days	14 days	21 days
SC-1	45 °C	8.45 (mm/day)	No growth	No growth
LE-1	38 °C	No growth	No growth	No growth

Study of lethal temperature

Along with the optimum growth rate at different temperatures, studies were undertaken to examine the maximum temperature tolerance limits and lethal temperature of both the wild strains and the results presented vide (Table 2a). The strain SC-1 was exposed to the maximum temperature of 45°C and later temperature was altered to the optimum range of 30°C after 7, 14 and 21 days of exposure period. The results clearly indicated that, the strain can tolerate the maximum temperature of 45°C up to 7 days and revived the growth to normal conditions (9 mm per day), whereas exposure period 14 days found to be lethal as there was no revival of growth when altered the temperature to 30°C. The upper limit of the temperature tolerance for LE-1 was 35°C for 12 days where the revival of growth (7.5 mm per day) was observed after exposing to the optimum temperature of 25°C. Whereas, the mycelium was completely killed at an exposure of 7 days at 38°C, where there was no revival of growth.

In conclusion, Experimental findings of comparative evaluation of culture media, pH and temperatures for mycelial growth of one each of wild strains of *Schizophyllum commune* and *Lentinula edodes* concluded that PDA medium with a pH range of 5.0 to 8.0 at a temperature of 30 °C and MEA medium with a pH range of 4.5 to 6.5 at a temperature of 25 °C gave the better growth of mycelium for *Schizophyllum commune* and *Lentinula edodes* respectively and appeared to better in maintenance of pure culture. This investigation will help for the selection of culture media, pH and optimum temperature for the maintenance of genetic materials.

References

- Arpita, M.T., and Bhupendra, N.T. 2013. Biochemical constituents of a wild strain of *Schizophyllum commune* isolated from Achanakmar-Amarkantak Biosphere Reserve (ABR), India. World J Microbiol Biotechnol 29, 1431-1442.

- Hem Lata, and Sharma, S.R. 2012. Evaluation of culture conditions for the vegetative growth of different strains of *Lentinula edodes*. *Mushroom Research* 21(1), 35-42.
- Martnexplores, H.E., Mayacorts, D.C., Figueroacardenas, J.D., Gamicaromo, M.G., and Poncesaavedra, J. 2009. Chemical composition and physicochemical properties of shiitake mushroom and high fibre products. *Journal of Food* 7(1), 7-14.
- Neha, J., and Bharat, P. 2012. Medicinal mushrooms: A blessing to the mankind. *Asian J of Research in Pharm Sci* 2(1), 12-15.
- Philippoussis, A.N., Diamantopoulou, P.A., and Zervakis, G.I. 2003. Correlation of the properties of several lignocellulosic substrates to the crop performance of the shiitake mushroom *Lentinula edodes*. *World J of Microb & Biotech* 19, 551-557.
- Sharma, S.R., Kumar, S., and Sharma, V.P. 2006. Physiological requirement for cultivation of Malaysian strain of shiitake, *Lentinula edodes*. *J Mycol Pl Pathol* 36(2), 149-152.
- Singh, S.K., Upadhyay, R.C., and Verma, R.N. 2000. Physico-chemical preferences for efficient mycelial colonization in edible mushrooms. *Mushroom Research* 9(2), 85-89.
- Tnglet, R.S., Song, M., Hensen, C.L., and Hwang, S. 2006. Cultivation of *Lentinula edodes* mycelia using whey permeate as an alternative growth substrate. *J Dairy Science* 89, 1113-1115.

How to cite this article:

Praveen Kumar Reddy, B., A. Uma Rajashekhar, P. Harikrishna and Lavanya, A.V.N. 2017. Cultural and Physiological Studies on Wild Mushroom Specimens of *Schizophyllum commune* and *Lentinula edodes*. *Int.J.Curr.Microbiol.App.Sci*. 6(7): 2352-2357.
doi: <https://doi.org/10.20546/ijcmas.2017.607.278>